

Phosphoric Acid

Livestock

Identification of Petitioned Substance

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2			
3	Chemical Names: phosphoric acid;	17	FSD-34™ (Diversey)
4	orthophosphoric acid (IUPAC name); H ₃ PO ₄	18	Demand low foaming anionic acid sanitizer
5		19	(Diversey)
6	Other Names: hydrogen phosphate;	20	Dividend anionic acid sanitizer (Diversey)
7	metaphosphoric acid; pyrophosphoric acid;	21	Hydri-San No. 468 (Hydrite)
8	white phosphoric acid; O-phosphoric acid;	22	
9	trihydroxidophosphorus; vococid;		CAS Numbers: 7664-38-2
10	orthophosphate; sonac; wc-reiniger;		
11	orthophosphoramidate		Other Codes:
12			UNII-E4GA8884NN
13	Trade Names:		E-number E338
14	HD CIP ACID™ (Aspen Veterinary Resources)		EC/EINECS 231-633-2
15	Acid Clean (Astro Products, Inc.)		EPA Pesticide Chemical Code 076001
16	Agrosan plus Acid Sanitizer (AgroChem)		
23			

Summary of Petitioned Use

Phosphoric acid is currently listed as an allowed substance in organic livestock production for use as a disinfectant, sanitizer, or cleaner for equipment.

§ 205.603 Synthetic substances allowed for use in organic livestock production.

In accordance with restrictions specified in this section the following synthetic substances may be used in organic livestock production:

(a) As disinfectants, sanitizer, and medical treatments as applicable.

(25) Phosphoric acid - allowed as an equipment cleaner, *Provided*, That, no direct contact with organically managed livestock or land occurs.

This limited scope technical report serves to support the sunset review of phosphoric acid in organic livestock production. The National Organic Standards Board (NOSB) Livestock Subcommittee has requested answers to two questions from the technical report template. These two questions serve as the focus of this limited report:

Evaluation Question #11: Describe all natural (non-synthetic) substances or products which may be used in place of a petitioned substance (7 U.S.C. § 6517 (c) (1) (A) (ii)). Provide a list of allowed substances that may be used in place of the petitioned substance (7 U.S.C. § 6518 (m) (6)); and

Evaluation Question #12: Describe any alternative practices that would make the use of the petitioned substance unnecessary (7 U.S.C. § 6518 (m) (6)).

Characterization of Petitioned Substance

Mechanical removal of organic residues is an important first step in cleaning livestock facilities and equipment. Subsequent disinfecting of equipment and livestock facilities is an essential second step in disease prevention and control. In order for disinfectants to be effective, attached organic material and

57 mineral scale need to be removed. Mineral scale can result when hard water is used in production settings.
58 It is typically the combination of calcium and magnesium compounds that precipitate out of water and
59 collect on surfaces. Water hardness will affect the likelihood and quantity of mineral scale deposition. High
60 levels of calcium, magnesium, and alkalinity¹ are all components that increase the potential for scale
61 formation (Sengupta, 2013).

62
63 In livestock facilities, phosphoric acid is used in both Clean-In-Place (CIP) and non-CIP systems² to remove
64 encrusted surface matter and mineral scale found on metal equipment. The chemical reaction of the acid
65 with minerals found in deposits makes them water soluble and thus easier to remove. For cleaning
66 purposes, phosphoric acid is often combined with a surfactant, usually a detergent. An example is
67 dodecylbenzene sulfonic acid (DDBSA) which is a component of the commercial product Hydri-san. It is
68 also in StarSan, a steel and glass sanitizing product commonly used in the beverage industry.

69
70 Phosphoric acid is sometimes used to remove resistant biofilms, colonies of microorganisms that attach to a
71 surface and are protected by a self-generated protective film of polysaccharide (Muhammad et al., 2020).
72 Surfaces covered with mineral scale are particularly susceptible to biofilm attachment. It is important to
73 note that when mineral scale is dislodged, the biofilm is also dislodged. Smooth surfaces are more difficult
74 to colonize. Research indicates that biofilm bacteria are up to 1000-times more resistant to disinfectants
75 than non-biofilm forming bacteria (Oliveira, 2014).

76
77 **Evaluation Question #11: Describe all natural (non-synthetic) substances or products which may be**
78 **used in place of a petitioned substance (7 U.S.C. § 6517 (c) (1) (A) (ii)). Provide a list of allowed**
79 **substances that may be used in place of the petitioned substance (7 U.S.C. § 6518 (m) (6)).**

80
81 *Non-synthetic alternatives*
82 No non-synthetic alternatives effective at removing encrusted surface matter and mineral scale were found.
83 Previous USDA technical reports for phosphoric acid (USDA, 2003; USDA, 2021) suggested a review of
84 scouring compounds and enzymatic cleaners as potential alternatives to phosphoric acid.

85
86 Scouring compounds are also known as chemical abrasives. They are normally manufactured from inert or
87 mildly alkaline materials and are typically combined with various soaps (Marriott et al., 2018). They are
88 then used with brushes or metal sponges. Neutral scouring compounds can be combined with acid
89 cleaners for removal of alkaline deposits and encrusted materials. Neutral scouring compounds are made
90 from such items as volcanic ash, pumice, silica flours, and feldspar. They are used in manual scrubbing and
91 scouring procedures. Slightly alkaline scouring compounds include borax and sodium bicarbonate. No
92 published research could be found on the use of such compounds in animal facilities.

93
94 Enzymatic cleaners are now available on the market but are marketed in industries other than organic
95 agriculture. Enzymes are proteins which catalyze chemical reactions. They break down soils and stains,
96 and they are typically mild, noncorrosive, and safe to handle. The main industry using enzymes for
97 cleaning is the clothing industry, where enzymes enhance biofilm removal (Stiefel et al., 2016).

98
99 It is now known that 40-80% of bacterial cells are able to form biofilms (Flemming and Wuertz, 2019).
100 Biofilms are complex surface-attached communities of microorganisms. These communities can consist of
101 single microbial species or a combination of species of bacteria, protozoa, archaea (single-celled,
102 prokaryotic microorganisms that includes methanogens and those of harsh environments), algae,
103 filamentous fungi, and yeast. They strongly attach to each other and to biotic or abiotic surfaces. Biofilms
104 can result in disease outbreaks. For example, biofilm formation was found to make major mastitis-causing
105 bacteria more resistant to the disinfectants typically used on commercial dairy-farms (Tremblay et al.,
106 2014). Other problems caused by biofilms include food spoilage, pipe fouling, and ship hull fouling.

¹ Alkalinity refers to a water source's ability to neutralize acidity. Carbonate, bicarbonate, hydroxide, borate, silicate, and phosphate contribute to alkalinity (Sengupta, 2013).

² Clean-in-Place refers to cleaning the interior surfaces of pipes and equipment without dismantling them first. Non-CIP would involve at least some dismantling of the equipment before cleaning.

107
108 Non-synthetic alternatives to manage bacterial populations are currently under development. One solution
109 is referred to as 'positive biofilms.' Positive biofilms involve using beneficial bacteria that are able to form
110 biofilms that outcompete undesirable microorganisms (Guéneau et al., 2022). Such products would be
111 applied to building surfaces to guide the microbial ecology of biofilms after cleaning and disinfection
112 procedures. The positive biofilms limit the proliferation of undesirable microorganisms, such as *Salmonella*
113 spp., *Escherichia coli*, *Enterococcus faecalis*, and *Enterococcus cocorum*, in building through nutritional and
114 spatial competition. Most commercial 'positive biofilm' products are composed of species such as
115 *Lactococcus* spp., *Lactobacillus* spp., or *Pediococcus* spp., often in combination with *Bacillus* spp. Large-scale
116 evaluation of commercial products is still being conducted. *Bacillus subtilis* is an industrially important
117 bacterium that forms rough biofilms at the air-liquid interface instead of on the surface of a solid phase in a
118 liquid (Morikawa, 2006). This permits the control of infection caused by plant pathogens and the reduction
119 of steel corrosion. It also allows for the exploration of novel compounds that could be used to control
120 harmful biofilm formation.

121
122 Lu et al. (2019) reviewed several natural products as potential anti-biofilm agents, including anti-biofilm
123 therapeutics undergoing clinical trials. There are anti-biofilm agents extracted from medicinal plants such
124 as garlic; *Cocculus trilobus*; *Coptis chinensis*; cranberry polyphenols; *Herba patriniae*; *Ginkgo biloba*; phloretin,
125 which is abundant in apples; citrus limonoids; and quercetin which exists in many fruits, vegetables, and
126 grains. There are currently no commercial products available that are made from these substances.

127
128 New mixtures of products may prove beneficial as well. Rocha e Silva et al. (2020) described a product
129 composed of a natural solvent (cottonseed oil), a plant-based surfactant agent (saponin), and two natural
130 stabilizers (carboxymethylcellulose and glycerine). The authors reported that the formulation was stable,
131 nontoxic, and highly efficient. It removed 100% of heavy oil from glass and metallic surfaces. Similar
132 products could be developed for cleaning animal housing and equipment.

133
134 It is important to look at other industries which also depend on cleaning for potential solutions to problems
135 encountered in cleaning and disinfecting livestock housing and equipment. For example, biocleaning has
136 been developed for artifact restoration (Martino et al., 2020). The products used include sulfate-reducing
137 bacteria (e.g., *Pseudomonas stutzeri*) and hydrolytic enzymes.

138
139 *Synthetic Alternatives*

140 Earlier USDA technical reports (USDA 2003; 2021) on phosphoric acid have indicated that other strong
141 acids have been used for cleaning operations but are not as effective or practical as phosphoric acid. These
142 alternative acids included hydrochloric, hydrofluoric, sulfamic, sulfuric, and nitric acids. Acids have the
143 ability to dislodge and dissolve mineral scale as long as the strength of the acid is high enough. When they
144 dislodge the mineral scale, they also dislodge the biofilm and are thus important in its removal. Nitric and
145 sulfuric acids are too corrosive to metal to be practical in livestock operations. Hydrochloric acid is used in
146 descaling metals but is a health hazard because of toxic hydrogen chloride gas. Additionally, none of these
147 acids besides phosphoric is currently permitted in organic livestock production. The previous technical
148 reports concluded that phosphoric acid was more practical than these other alternative acids since it is the
149 lowest in corrosiveness and is compatible with many surfactants.

150
151 Peracetic acid (CAS #-79-21-0) is permitted on the National List at § 205.603(a)(24) for sanitizing organic
152 livestock facilities and processing equipment. The 2016 USDA technical report on peracetic acid (USDA,
153 2016) indicates that it is also effective in removing biofilms. Oxalic acid also effectively removes iron oxide
154 rust without attacking the metal, but is currently only permitted in organic apiculture. Precautionary steps
155 are also necessary with oxalic acid, since it reacts with hard-water constituents and forms a poisonous
156 precipitate, calcium oxalate (Marriott et al., 2018).

157
158 Previous USDA technical reports for phosphoric acid used in organic processing/handling (USDA, 2003;
159 USDA, 2021) suggested a review of colloids, sequestrants, and auxiliary compounds used in cleaners as
160 potential alternatives to phosphoric acid. However, specific materials within each of these classes of

161 products would need to be petitioned for addition to the National List and evaluated individually. Further,
162 combinations of materials found to have multiple beneficial properties for cleaning and sanitizing
163 (Shkromada et al. 2021) would also need to have any synthetic components assessed for compliance with
164 the National List.

165
166 Electrolyzed oxidized water (EOW) is a relatively new material that can be used for disinfecting animal
167 facilities (Hao et al., 2013a, b; Rahman and Murshed, 2019). EOW is on the National List under
168 §205.603(a)(10)(iv), Chlorine Materials, Hypochlorous acid – generated from electrolyzed water. EOW is
169 produced on-site by electrolysis of a 0.1% sodium chloride solution in an electrolysis chamber. The anode
170 (positive electrode) and the cathode (negative electrode) of the chamber are separated by a diaphragm to
171 form two separate compartments. The anode acidic EOW has a low pH (typically 2.3-3.0), a high oxidation-
172 reduction potential (ORP) (>1000 mV), and contains relative concentrations of chlorine, hypochlorous acid,
173 and hypochlorite (Fenner, 2005). The cathode alkaline EOW has a high pH and a low ORP. The physical
174 properties and chemical composition of the EOW will vary depending on the concentration of sodium
175 chloride, the amperage used, the electrolysis time, and the flow rate of the water (Fenner, 2005).

176
177 Anode EOW was shown to have strong anti-microbicidal activity against a broad variety of bacterial
178 pathogens. There were, however, marked differences in the sensitivity of the different bacterial strains
179 tested, with the gram-negative bacterium *Proteus mirabilis* and the gram-positive bacterium *Salmonella*
180 *aureus* being more susceptible than the gram-negative *Pseudomonas aeruginosa* and gram-positive
181 *Enterococcus faecium*. The latter species required more exposure time in order to be killed. EOW has also
182 been shown to be effective on mycobacteria (e.g., *Mycobacterium avium*) as well as bacterial endospores. It is
183 also reported to have fungicidal activity and can inactivate bacterial and fungal toxins (Fenner, 2005).

184
185 **Evaluation Question #12: Describe any alternative practices that would make the use of the petitioned**
186 **substance unnecessary (7 U.S.C. § 6518 (m) (6)).**

187
188 Cleaning and disinfecting (C&D) are essential elements in disease prevention and control on any animal
189 production or processing facility. In fact, Schmidt (1997) discusses the properties of the types of
190 contaminants that can be found on surfaces (fat-based, protein-based, etc.) and how these properties,
191 together with the properties of the different types of surfaces, will affect the C&D procedures. This would
192 include choice of cleaning agents. However, C&D is much easier when there is no encrusted material or
193 mineral scale to remove. Preventing such occurrences, therefore, would be important in reducing or
194 eliminating the need for phosphoric acid as a cleaning tool.

195
196 The ability of materials to encrust a surface depends on the surface tension of those materials (Marriott et
197 al., 2018). The type of material used will also affect the type of C&D chemicals that can be used. Marriott et
198 al. (2018) characterized the various surfaces with regard to their suitability.

- 199 • Black metals are prone to rust so are often tinned or galvanized. Neutral detergents are
200 recommended for cleaning such surfaces.
- 201 • Tin surfaces are easily corroded by strong alkaline and acid cleaners.
- 202 • Cement should be dense, acid resistant, and non-dusting. Acid brick may be used in place of
203 concrete.
- 204 • Glass should be smooth and impervious and should be cleaned with moderately alkaline or
205 neutral detergents.
- 206 • Rubber should be nonporous, non-spongy, and not affected by alkaline detergents. All rubber
207 surfaces can be impacted by organic solvents and strong acids.
- 208 • Stainless steel is generally resistant to corrosion. It has a smooth surface and is impervious.

209
210 Microbial populations may form biofilms as a protective response to environmental stresses such as
211 UV radiation, desiccation, limited nutrients, high pressure, and antimicrobial agents. The events leading to
212 biofilm formation are complex but are believed to start with reversible attachment. Both inert and
213 biological surfaces can be used for the initial bacterial attachment. The physicochemical properties of the
214 surface will determine how quickly biofilms develop. These properties include surface roughness,

215 hydrophobicity, surface charge, and presence of conditioning films. The choice of surfaces in animal
216 housing, therefore, can affect biofilm formation. Remodeling the surface or coating the surface with
217 substances that do not encourage bacterial adhesion are strategies that could be implemented to impede
218 the establishment of bacterial biofilms (Flemming and Wuertz, 2019).

219
220 The chemical properties of the water used in cleaning operations should be considered. Hard water with
221 varying amounts of calcium, magnesium, and other alkali metals interferes with the effectiveness of
222 cleaning products. Hard water also contributes to the formation of precipitates. Such precipitates allow for
223 the accumulation of debris and microorganisms, making effective C&D difficult. If a farm has hard water,
224 it may be more economical to use a water softener to mitigate the problem of precipitates, rather than
225 relying on more C&D (Marriott et al. 2018).

226
227 The removal of encrusted material from a surface can be done through mechanical action of high-pressure
228 water, steam, air, and scrubbing. It can also be done through the use of surfactants that reduce surface
229 tension of the cleaning medium and allow more close contact with the material.

230
231 The Federal Pasteurized Milk Ordinance (PMO) requires milking operations to employ effective C&D
232 procedures for product-contact surfaces of multi-use containers, utensils and equipment used in the
233 transportation, processing, condensing, drying packaging, handling and storage of milk or milk products
234 before each use (or at regular intervals for certain systems) (FDA, 2019). While the PMO does not mandate
235 specific materials for disinfection, it does reference food-contact surface sanitizing materials with tolerance
236 exemptions at 21 CFR 180.940. In addition to chemical sanitization, other approved methods include the
237 use of steam or hot water. Some methods maybe be recommended for certain applications over others. For
238 example, caustic solutions of sodium hydroxide, another material permitted on the National List at §205.
239 605(b), are recommended for soaker-type bottle washers, to be followed by clean water rinse or chemical
240 treatment to prevent recontamination (FDA, 2019).

241
242 Cleaning procedures in other industries may inform new and different means of preventing encrusted
243 material and mineral scale in livestock operations. The fouling of ship hulls is a significant obstacle to
244 efficient ship operation. Copper-based antifouling coatings have been used, however, bans are being
245 considered because of copper leaching into the water (Chambers et al., 2006). Holm et al. (2003) looked at
246 non-toxic alternatives to the toxic antifouling paints being used on ship hulls. One alternative was silicone
247 coatings. The authors then developed a portable rotating brush device that can be used to clean hulls
248 without damaging the coating. No published research looking at the use of this material in animal
249 agriculture facilities was found in the literature reviewed for this report. However, this alternative may be
250 worth investigating for the treatment of animal housing and equipment to minimize attachment of organic
251 material and mineral scale, and increase ease of cleaning.

252

Report Authorship

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255 The following individuals were involved in research, data collection, writing, editing, and/or final
256 approval of this report:

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264 All individuals are in compliance with Federal Acquisition Regulations (FAR) Subpart 3.11 – Preventing
265 Personal Conflicts of Interest for Contractor Employees Performing Acquisition Functions.

266

References

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269 Bidi, H., M.E. Touhami, Y. Baymou, I-M. Chung, H. Lgaz and S. Zehra. 2020. Toward the development of
270 an innovative descaling and corrosion inhibition solutions to protect mild steel equipment: an
271 experimental and theoretical approach. *Chemical Engineering Communications* 2087(5):632-651.
- 272 Chambers, L.D., K.R. Stokes, F.C. Walsh, and R.J.K. Wood. 2006. Modern approaches to marine antifouling
273 coatings. *Surface & Coatings Technology* 201:3642-3652.
- 274 Coughlan, L.M., P.D. Cotter, C. Hill, and A. Alvarez-Ordóñez. 2016. New weapons to fight old enemies:
275 Novel strategies for the (Bio)control of bacterial biofilms in the food industry. *Frontiers in Microbiology*,
276 Volume 7. Article 1641. Available online at
277 <https://www.frontiersin.org/articles/10.3389/fmicb.2016.01641/full>
- 278 FDA (U.S. Food and Drug Administration). 2019. Pasteurized Milk Ordinance. Available online at
279 <https://www.fda.gov/media/140394/download> (Accessed 10/7/2022).
- 280 Fenner, D.C. 2005. Thesis dissertation: Antimicrobial activity of electrolyzed oxidizing water using
281 standard in-vitro test procedures for the evaluation of chemical disinfectants.
- 282 Flemming, H.C. and S. Wuertz. 2019. Bacteria and archaea on Earth and their abundance in biofilms. *Nature*
283 *Reviews Microbiology* 17: 247-260,
- 284 Guéneau, V., J. Plateau-Gonthier, L. Arnaud, J-C. Piard. 2022. Positive biofilms to guide surface microbial
285 ecology in livestock buildings. *Biofilm* Volume 4. Available online at
286 <https://www.sciencedirect.com/science/article/pii/S2590207522000090>
- 287 Hao, X.X., B.M. Li, C.Y. Wang, Q. Zhang, and W. Cao. 2013a. Application of slightly acidic electrolyzed
288 water for inactivating microbes in a layer breeding house. *Poultry Science* 92:2560-2566.
- 289 Hao, X.X., B.M. Li, Q. Zhang, B.Zh. Lin, L.P. Ge, C.Y. Wang, and W. Cao. 2013b. Disinfection effectiveness
290 of slightly acidic electrolyzed water in swine barns. *Journal of Applied Microbiology* 115:703-710.
- 291 Holm, E.R., E.G. Haslbeck, and A.A. Horinek. 2003. Evaluation of brushes for removal of fouling from
292 fouling-releasing surfaces, using a hydraulic cleaning device. *Biofouling* 19(5):297-305.
- 293 Lu, L., W. Hu, Z. Tian, D. Yuan, G. Yi, Y. Zhou, Q. Cheng, J. Zhu, and M. Li. 2019. Developing natural
294 products as potential anti-biofilm agents. *Chinese Medicine* 14:11-28.
- 295 Marriott, N.G., M.W. Schilling, and R.B. Gravani. 2018. Cleaning compounds. In: *Principles of Food*
296 *Sanitation*. Food Science Text Series. Springer International Publishing AG. Pages 151-174.
- 297 Martino, M., A. Balloi, and F. Palla. 2020. Biocleaning. Chapter 4 in *Biotechnology and Conservation of Cultural*
298 *Heritage*, Second Edition. F. Palla and G. Barresi (eds). Pages 71-96.
- 299 Morikawa, M. 2006. Beneficial biofilm formation by industrial bacteria *Bacillus subtilis* and related species.
300 *Journal of Bioscience and Bioengineering*. 101(1):1-8.
- 301 Muhammad, M.H., A. L. Idris, X. Fan, Y. Guo, Y. Yu, X. Jin, J. Oiu, X. Guan, and T. Huang. 2020. Beyond
302 Risk: Bacterial biofilms and their regulating approaches. *Frontiers in Microbiology* 11: Article 928.
- 303 Oliveira, C.A.F. 2014. On the relevance of microbial biofilms for persistence of *Staphylococcus aureus* in
304 dairy farms. *Advances in Dairy Research* 2(2):1000e109.
- 305 Rahman, S.M.E. and H.M. Murshed. 2019. Chapter 8. Application of electrolyzed water on livestock. In:
306 *Electrolyzed Water in Food: Fundamentals and Applications*. T. Ding, D-H. Oh, and D. Liu (eds). Springer
307 Publishing. Pages 205-222.
- 308 Rocha e Silva, N.M.P., F.C.G. Almeida, and F.C.P. Rocha e Silva. 2020. Formulation of a biodegradable
309 detergent for cleaning oily residues generated during industrial processes. *Journal of surfactants and*
310 *detergents* 23(6):1111-1123.
- 311 Schmidt, R.H. 1997 (reviewed 2009). Basic elements of equipment cleaning and sanitizing in food
312 processing and handling operations. University of Florida Factsheet FS14/FS077. Available online at
313 <https://edis.ifas.ufl.edu/publication/FS077> (Accessed August 25, 2022).
- 314 Sengupta, P. 2013. Potential Health Impacts of Hard Water. *International Journal of Preventive Medicine*,
315 4(8), 866-875.
- 316 Shkromada, O., T. Fotina, R. Petrov, L. Nagorna, O. Bordun, M. Barun, O. Babenko, M. Karpulenko, T.
317 Tsarenko, and V. Solomon. 2021. Development of a method of protection of concrete floors of animal
318 buildings from corrosion at the expense of dry disinfectants. *Eastern-European Journal of Enterprise*
319 *Technologies* 4(6(112)):33-40.

- 320 Song, J., H. Ruan, L. Chen, Y. Jin, J. Zheng, R. Wu and D. Sun. 2021. Potential of bacteriophages as
321 disinfectants to control of *Staphylococcus aureus* biofilms. *BMC Microbiology* 21:57-74.
- 322 Stiefel, P., S. Mauerhofer, J. Schneider, K. Maniura-Weber, U. Rosenberg, and Q. Ren. 2016. Enzymes
323 enhance biofilm removal efficiency of cleaners. *Antimicrobial agents and Chemotherapy* 60(6):3647-3652.
- 324 Tremblay, Y.D.N., V. Caron, A. Blondeau, S. Messier, and M. Jacques. 2014. Biofilm formation by
325 coagulase-negative staphylococci: Impact on the efficacy of antimicrobials and disinfectants commonly
326 used on dairy farms. *Veterinary Microbiology* 172:511-518.
- 327 USDA (United States Department of Agriculture). 2003. Phosphoric acid technical evaluation report.
328 Available online at [https://www.ams.usda.gov/sites/default/files/media/Phos acid 2002 technical](https://www.ams.usda.gov/sites/default/files/media/Phos%20acid%202002%20technical%20advisory%20panel%20report.pdf)
329 [advisory panel report.pdf](https://www.ams.usda.gov/sites/default/files/media/Phos%20acid%202002%20technical%20advisory%20panel%20report.pdf) (Accessed August 30, 2022).
- 330 USDA (United States Department of Agriculture). 2016. Technical evaluation report: Peracetic acid –
331 Handling/Processing. Available online at
332 [https://www.ams.usda.gov/sites/default/files/media/Peracetic Acid TR 3_3 2016 Handling](https://www.ams.usda.gov/sites/default/files/media/Peracetic%20Acid%20TR%203_3_2016%20Handling%20final.pdf)
333 [final.pdf](https://www.ams.usda.gov/sites/default/files/media/Peracetic%20Acid%20TR%203_3_2016%20Handling%20final.pdf) (Accessed August 30, 2022).
- 334 USDA (United States Department of Agriculture). 2021. Phosphoric acid technical report. Available online
335 at <https://www.ams.usda.gov/sites/default/files/media/USDAHandlingPhosphoricAcid.pdf>
- 336 Van Immerseel, F., K. Luyckx, K. De Reu, and J. Dewulf. 2020. Chapter 6. Cleaning and disinfection. In
337 Biosecurity in Animal Production and Veterinary Medicine: From Principles to Practice (J. Dewulf and
338 F. Van Immerseel, editors). ProQuest Ebook.