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Document Cover Sheet

<https://www.ams.usda.gov/rules-regulations/organic/national-list/petitioned>

Document Type:

**National List Petition or Petition Update**

A petition is a request to amend the USDA National Organic Program's National List of Allowed and Prohibited Substances (National List).

Any person may submit a petition to have a substance evaluated by the National Organic Standards Board (7 CFR 205.607(a)).

Guidelines for submitting a petition are available in the NOP Handbook as NOP 3011, National List Petition Guidelines.

Petitions are posted for the public on the NOP website for Petitioned Substances.

**Technical Report**

A technical report is developed in response to a petition to amend the National List. Reports are also developed to assist in the review of substances that are already on the National List.

Technical reports are completed by third-party contractors and are available to the public on the NOP website for Petitioned Substances.

Contractor names and dates completed are available in the report.

# Lignins

## Crops

### Identification of Petitioned Substance

**Chemical Names:**

Soda lignin; Sodium lignin; Sodium carbonate lignin; Kraft lignin; Lignin sulfonate

**CAS Numbers:**

8068-05-1 Alkali lignin, alkali soluble lignin, kraft lignin, soda lignin  
 9005-53-2 Lignin  
 8062-15-5 Lignin sulfonate  
 8061-51-6 Sodium lignin sulfonate  
 8061-54-9 Magnesium lignin sulfonate  
 8061-53-8 Ammonium lignin sulfonate  
 8061-52-7 Calcium lignin sulfonate

**Other Names:**

Alkali lignin; Alkaline lignin; Sulfur-free lignin; Sulfur-free anionic sodium carbonate lignin

**Trade Names:**

Polybind 300

**Other Codes:**

N/A

### Summary of Petitioned Use

Sodium carbonate lignin has been petitioned for use in organic crop production through addition to the National List at 205.601(j)(4) of “lignins” as a dust suppressant. Lignin sulfonate, another type of lignin, is currently listed at 205.601(j)(4) for use as a chelating agent and dust suppressant.

### Characterization of Petitioned Substance

This report focuses primarily on alkali lignins, including sodium carbonate lignin. Supplemental information on other lignin derivatives from the kraft (sulfonated), sulfite, and organosolv processes is also included but is not the primary focus of this report. Lignins that are chemically modified subsequent to or beyond the pulping and its associated extraction steps are outside of the scope of this report. Table 1 provides a summary of the lignin derivatives covered in this report.

**Table 1. Lignin derivatives addressed in this report**

	Lignin type			
	Soda lignin	Organosolv lignin	Kraft lignin	Lignin sulfonate
<b>Alternate or additional name(s)</b>	Alkali lignin; Alkaline lignin; Sodium lignin; Sodium carbonate lignin	Acetosolv lignin; Organocell lignin	Alkali lignin; Alkaline lignin; Alkaline sulfite lignin; Lignin sulfonate	Lignin sulfonate; Salts of lignosulfonic acid
<b>Source process</b>	Soda pulping	Organosolv pulping	Kraft pulping	Sulfite pulping
<b>Sulfonated end product</b>	No	No	Extent of sulfonation depends on process	Yes

**Composition of the Substance:**

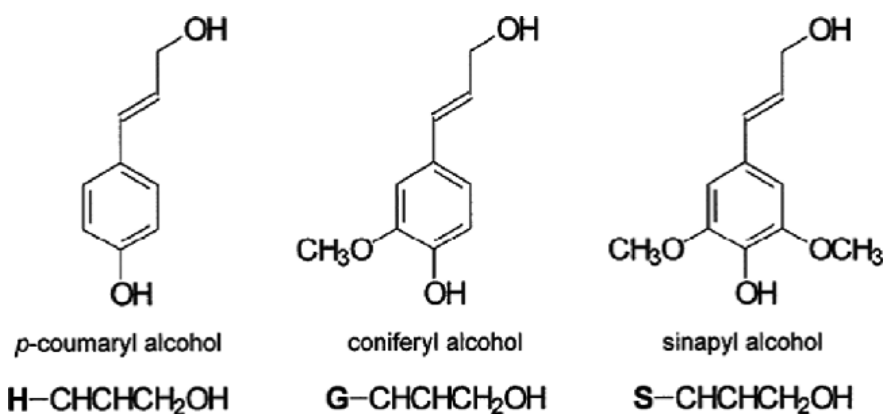
Native lignins found in plant cell walls are amorphous, complex biopolymers (Peretti, Barton and Teixeira Mendonca 2016). They are made up of phenylpropane units (C6-C3), known as monolignols. These monolignols are *p*-coumaryl, coniferyl, and sinapyl alcohols. The main difference between these different phenylpropane

37 derivatives is the number of methoxyl groups attached to the phenolic unit (Peretti, Barton and Teixeira  
 38 Mendonca 2016) (see Figure 1). There are other, less abundant lignin monomers of different phenylpropane units  
 39 and some carbohydrate moieties. In native lignin biosynthesis, all of these components combine in various ways  
 40 to form three-dimensional, cross-linked polymers that do not have one regular, ordered macromolecule. Thus,  
 41 lignins are physically and chemically heterogeneous, lacking a defined primary structure (Calvo-Flores, et al.  
 42 2015; Košíková and Gregorová 2005; Karak 2016; Du, et al. 2013; Peretti, Barton and Teixeira Mendonca 2016;  
 43 Garcia-Valls and Hatton 2003).

44

45 **Figure 1. Building blocks of lignin. (Du, et al. 2013).** The subindices H, G, and S indicate the alcohol  
 46 moiety of each monolignol: *p*-hydroxyphenyl, guaiacyl, and syringyl, respectively.

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50 Industrial processes such as paper pulping and bioethanol production separate different components of  
 51 plant biomass and produce lignins as a byproduct, which may then be commercialized. These lignin  
 52 byproducts differ from native lignin in size and structure (Ahvazi and Ngo 2018) depending on the method  
 53 of extraction used (Agrawal, Kaushik and Biswas 2014). The chemical structure of these lignin polymers  
 54 includes significant quantities of three important functional groups: phenolic hydroxyl groups, aliphatic  
 55 hydroxyl groups, and carboxylic acid groups (Ahvazi and Ngo 2018). The types and quantities of these  
 56 functional groups in any lignin byproduct depend largely on the pulping process used to extract the  
 57 byproduct. These extracted lignins also differ in size of polymeric fragments depending on the methods  
 58 used to recover them from the pulping black liquor (i.e., the extraction liquid resulting from the pulping  
 59 process) (Ahvazi and Ngo 2018).

60

61 The petitioned material results from a paper pulping process that uses sodium carbonate and sodium  
 62 hydroxide to extract lignin. In contrast, lignin sulfonates are produced using a sulfite chemical pulping  
 63 process, where sulfur dioxide is used to extract lignin under pressure. A 2011 technical report on lignin  
 64 sulfonate includes detailed information on this particular type of lignin (USDA NOP 2011). The petitioner  
 65 of the present substance states that its composition is made up of lignin fragments, carbohydrates from the  
 66 breakdown of hemicellulose, and residual sodium carbonate. The lignin in this mixture is in varying states  
 67 of degradation and complexation with cellulose and hemicellulose (Legnochem 2019). Soda lignin has little  
 68 to no sulfur and a low quantity of hemicellulose but can contain high amounts of silicate and nitrogen  
 69 depending on the extraction procedure used (Espinoza-Acosta et. al. 2016). Lignins from soda ash pulping  
 70 have also been observed to have higher sodium ion content than other lignins (Vivekanandhan, Misra and  
 71 Mohanty 2015). One report on sulfur-free lignin suggested that these lignin derivatives contain more lignin  
 72 with macromolecular size (about 50–75 percent) and structure akin to native lignin than those in lignin  
 73 sulfonates (Liu, et al. 2020). The sulfur-free lignin used in the study was a byproduct of ethanol production  
 74 and contained 91.2 percent lignin, 0.12 percent residual cellulose/hemicellulose, and 0.67 percent ash (Liu,  
 75 et al. 2020). The sodium content of an alkali kraft lignin precipitated from the pulping black liquor using  
 76 carbon dioxide was reported to be 10 kg, or 1 percent, of sodium per ton of lignin (Stigsson and Lindstrom  
 77 2007).

78

79 Because the methods used to delignify plant matter influence the structure and size of the final lignin  
 80 byproduct, lignins are typically characterized by their extraction process (Serrano, et al. 2012). For example,

81 *soda lignins* are derived from cooking plant matter in sodium hydroxide. *Organosolv lignins* are obtained  
82 from the use of organic solvents used in the organosolv process. *Kraft lignins* are derived from the kraft  
83 process, and *lignin sulfonates* result principally from the sulfite pulping process. However, there are some  
84 areas of discrepancy in how lignins are reported in the literature, with inconsistent use of terms. For  
85 example, some reports refer to alkali or alkaline lignins as those resulting from the soda or soda ash process  
86 and as being distinct from kraft lignins (Marin, et al. 2017; Calvo-Flores, et al. 2015). Conversely, the kraft  
87 process uses the alkaline materials sodium hydroxide and sodium sulfide, and Collins (2019) lists alkali  
88 pulping as also being known as kraft pulping. Ahvazi and Ngo (2018) refer to “alkaline kraft extraction.”  
89 Another report refers to the kraft process as the “alkaline sulfite method” (Khrstova, et al. 2002). In some  
90 reports the term “alkali lignin” is not defined in terms of specific processing chemicals (Agrawal, Kaushik  
91 and Biswas 2014). No studies reviewed for this report expressly referred to lignin sulfonates as alkali  
92 lignins, though lignin sulfonates are also produced under strong alkali conditions. See Table 1 for alternate  
93 names associated with the various pulping processes and Table 3 for process details.

94  
95 Inconsistent terminology also appears in the literature when the sulfur content of lignin products is  
96 discussed. Numerous reports identify “sulfur-free lignin,” which may be derived from wood prehydrolysis  
97 (Košíková and Gregorová 2005), bioethanol production (Liu, et al. 2020), organosolv, soda (Collins 2019), or  
98 soda ash processes. Calvo-Flores et al. (2015) identify sulfur-free lignin as that obtained from soda,  
99 organosolv, and ionic liquid processes. Other lignins are acknowledged to contain sulfur, such as lignin  
100 sulfonates. Espinoza-Acosta et al. (2016) group kraft lignins with lignin sulfonates as sulfur-containing.  
101 However, Agrawal et al. (2014) note the availability of a commercial sulfur-free kraft lignin.

102  
103 Ahvazi and Ngo (2018) measured the sulfur content of various lignins, which differs primarily based on  
104 extraction method. They found that samples of soda lignin from wheat straw and organosolv lignin from  
105 hardwood both had <0.50 percent sulfur, while another soda lignin from wheat straw had a sulfur content  
106 of 1.19 percent. Two kraft lignins from hardwood and softwood showed sulfur values of 1.93 percent and  
107 1.32 percent, respectively. These values are in contrast to the 4.16 percent sulfur measured in a low-  
108 sulfonated kraft lignin from softwood and the 6.63 percent and 6.59 percent sulfur content found in lignins  
109 from sulfite pulping of softwood (Ahvazi and Ngo 2018).

110  
111 Calvo-Flores et al. (2015) similarly reported a sulfur content of 6.5 percent for lignin sulfonate compared to  
112 1.6 percent for kraft lignin. Thus, kraft lignin may or may not contain significant amounts of sulfur  
113 depending on its processing. Other lignins are identified as “sulfur-free lignins” without disclosing their  
114 processing chemicals (Sahoo, et al. 2011). Thus, identification of lignins by sulfur content does not directly  
115 correspond to the general process-based characterizations in every case.

116  
117 Although the extraction process is the predominant identifier for a lignin, the source plant material also has  
118 bearing on the makeup of the final lignin product. Lignins may be derived from hardwoods, softwoods, or  
119 herbaceous plants (Agrawal, Kaushik and Biswas 2014). Softwood lignins contain almost exclusively  
120 guaiacyl units, while hardwoods contain guaiacyl and syringyl units in different ratios (Ahvazi and Ngo  
121 2018), and herbaceous plants contain all of the three moieties (Ponnusamy, et al. 2019) with significant  
122 amounts of *p*-hydroxyphenyl units (Agrawal, Kaushik and Biswas 2014). Regardless of the plant source,  
123 however, all lignins have  $\beta$ -O-4 linkages as major interunit bonds (Calvo-Flores, et al. 2015).

#### 124 **Source or Origin of the Substance:**

125  
126 Lignin is a basic, albeit heterogeneous and complex, structural component of plants. It makes up  
127 approximately 20–30 percent of plant cell walls (Peretti, Barton and Teixeira Mendonca 2016), 30 percent of  
128 the total mass in softwoods, 20–25 percent of the total mass in hardwoods, and smaller percentages of the  
129 total mass in herbaceous species (Calvo-Flores, et al. 2015). Commercial lignins are obtained as byproducts  
130 of the paper pulping and bioethanol industries.

131  
132 The particular lignin derivative identified in the petition, sodium carbonate lignin, comes from soda ash  
133 pulping of wood chips, in which sodium carbonate and sodium hydroxide are used for extraction  
134 (Legnochem 2019). The milder reagents of sodium hydroxide and sodium carbonate – along with others  
135 such as aqueous ammonia mixed with potassium hydroxide, and sodium hydroxide with the pulping

136 additive anthraquinone (AQ) – have been reported to be more suitable for pulping straw (Marin, et al.  
137 2017) than stronger chemicals used in other processes.

138  
139 Based on existing literature, soda ash pulping is not in widespread use. Soda pulping is more common but  
140 is not the predominant pulping process. In the United States, soda pulping fell from 420,000 metric tons  
141 (MT) in 1960 to 300,000 MT in 1990. By comparison, kraft pulping in the United States produced 14,590 MT  
142 in 1960 and 50,000,000 MT in 1990 (Biermann 1996). Global kraft pulp production was approximately  
143 118,000,000 MT in 2015, accounting for 80% of global paper production (Fernández-Rodríguez, et al. 2019).  
144 The literature reviewed for this report indicates that sources of sodium carbonate lignin specifically may be  
145 limited, though there are numerous kinds of technical lignins that vary based on both their plant source  
146 and the method used to extract them.

147  
148 **Properties of the Substance:**

149 Lignin byproducts from plant processing differ from native lignin in terms of size and structure. The  
150 properties of these lignins also vary based on the origins of varied molecular and structural composition as  
151 well as different methods of extraction and recovery (Košíková and Gregorová 2005; Espinoza-Acosta, et al.  
152 2016). Ahvazi and Ngo (2018) report that the nature of a lignin's hydrophilic functional groups is  
153 dependent on the pulping process, and that the size of the polymeric pieces depend on the methods used  
154 to recover the lignins from the pulping black liquor. Both of these factors influence a lignin's properties.

155  
156 Espinoza-Acosta et al. (2016) conducted an assessment of molecular weight and polydispersity of different  
157 types of lignins, which is summarized in Table 2 below. Among the different lignins, lignin sulfonates have  
158 the highest overall molecular weight. However, determination of any particular lignin's molecular weight  
159 may vary depending on the measurement method used. For example, when measured as a solution  
160 property, the molecular weight will reflect the portion soluble in the organic solvent, whereas the  
161 molecular weight of the whole isolated lignin may be orders of magnitude greater (Calvo-Flores, et al.  
162 2015). A property perhaps more descriptive of the reactive potential of any given lignin is that of  
163 polydispersity. The polydispersity index (PDI) is a measure of the breadth of a polymer's molecular weight  
164 distribution. It is defined as  $M_w/M_n$ , where  $M_w$  is the average molecular weight and  $M_n$  is the number  
165 average molecular weight. A polymer with uniform molecular weight distribution for all of its polymeric  
166 components would have a PDI value of 1. Lignins have high polydispersity, meaning their polymeric units  
167 are diverse sizes.

168  
169 **Table 2. Physiochemical Properties of Technical Lignins. Adapted from Espinoza-Acosta, et al. 2016 .**

	Kraft lignins	Lignosulfonates	Soda lignins	Organosolv lignins
Molecular weight ( $M_w$ )	100 to 3000	20,000 to 50,000	800 to 3,000	500 to 4000
Polydispersity ( $M_w/M_n$ )	2.5 to 3.5	6 to 8	2.5 to 3.5	1.3 to 4.0
Impurities	Sulfur	Sulfur	Carbohydrates and ash	Carbohydrates and ash

170  
171 Vishtal and Kraslawski (2011) also summarized properties of different lignins and suggested wider ranges  
172 for the molecular weight and polydispersity of lignin sulfonates and higher molecular weight for kraft  
173 lignin than the values reported by Espinoza-Acosta, et al. (2016).

174  
175 The molecular mass of lignins varies based on nature of the raw material and the extraction method  
176 (Serrano, et al. 2012) and can also vary based on the pH and temperature applied during recovery (Peretti,  
177 Barton and Teixeira Mendonca 2016). Native lignin has high molecular weight and a condensed structure,  
178 which make it resistant to chemical and biological processing. Organosolv lignin has a lower molecular  
179 weight than soda lignin (Peretti, Barton and Teixeira Mendonca 2016) (Serrano, et al. 2012) because lignins  
180 in the soda process are so highly fractionated under the alkaline conditions that they repolymerize into  
181 larger fragments (Serrano, et al. 2012).

182  
183 Lignins can form many bonds. In paper pulp, lignin impedes the formation of bonding between the  
184 cellulose fibers (Marin, et al. 2017), which is why pulping processes aim to remove lignin. As aromatic  
185 polymers, lignins have many aromatic rings with hydroxyl and methoxyl functional groups that can act as  
186 hydrogen acceptors. They also have antioxidant properties (Espinoza-Acosta, et al. 2016). Lignins are  
187 readily oxidized, and readily condensed with phenols or thiols (Calvo-Flores, et al. 2015). Organosolv  
188 lignins are reported to contain more oxidized functionalities than alkali lignins (Peretti, Barton and Teixeira  
189 Mendonca 2016). The colloidal nature of lignins also makes it difficult to separate lignin precipitate from  
190 the extracting liquor (Hellstén, et al. 2013). Lignins are also not optically active (Calvo-Flores, et al. 2015).

191  
192 Lignin's chemical network of strong intra- and inter-molecular hydrogen bonds enable it to display  
193 thermoplastic behavior; however, upon initial application of high temperature (approximately 100 °C),  
194 lignin will behave as a thermoset material, hardening due to extensive cross-linking reactions (Collins  
195 2019). This would occur at much higher temperatures than would be naturally found in soils and thus  
196 would not be a concern for the health of the soil. One study comparing organosolv and soda lignins  
197 observed that soda lignins were the most thermally stable of the two (Serrano, et al. 2012).

198  
199 Soda lignin displays properties similar to those of kraft lignin, with the notable difference that it contains  
200 little to no sulfur (Košíková and Gregorová 2005; Wörmeyer et al. 2011; Laurichesse and Averous 2014).

#### 201 202 **Specific Uses of the Substance:**

203 Lignins have a wide range of agricultural and non-agricultural uses. They are abundant, variable, and  
204 naturally reactive due to their many different functional groups (Serrano, et al. 2012). In particular, the use  
205 of lignin sulfonates as binders, dust suppressants, and dispersing agents is well documented (Strassberger,  
206 Tanase and Rothenberg 2014).

#### 207 208 *Lignins used for binding*

209 Lignins are added in fertilizer formulations to improve granule formation by binding extrusion granules.  
210 This also improves the suspension of granules and the dispersion and suspension of concentrates and  
211 particles in fertilizer solutions. The addition of lignins also reduces fertilizer caking during storage and  
212 protects fertilizer components from UV photodegradation (Calvo-Flores, et al. 2015). Lignin polymers are  
213 employed in controlled-release fertilizer and pesticide formulations (Peretti, Barton and Teixeira Mendonca  
214 2016). Sodium carbonate lignin is referenced in several patents as one option for a binder in fertilizer  
215 chemical carrier substrates, along with lignin sulfonates, cane molasses, beet syrup, starch and starch  
216 derivatives, or any number of other potential binders (Welshimer, Dunn and Birthisel 2003; Ginn and Gray  
217 2018). In livestock production, lignins are used as pelletizing binders for animal feed to facilitate handling  
218 and preservation (Calvo-Flores, et al. 2015).

#### 219 220 *Lignins used for dust suppression*

221 Lignins are useful as binders in fertilizer and feed formulations because improving the quality of the  
222 pelletizing or granulation of these materials is a well-known measure for dust suppression (Kucera and  
223 Sawyer 2015). Fertilizers often contain an undesirable level of fine particles that can become airborne dust  
224 during the manufacture, storage, and transportation of the fertilizer. Dust can become airborne when the  
225 fertilizer materials undergo mechanical abrasion, when chemical reactions or curing processes continue  
226 after initial particle formation, when materials lose moisture during storage, or are otherwise affected by  
227 temperature and humidity conditions during handling and storage. Adding binders such as lignin  
228 increases fertilizer granule strength, thereby reducing the formation of dust during these activities (Kucera  
229 and Sawyer 2015).

230  
231 Lignin sulfonate is used for dust control on roads and for this application would be sprayed on the soil  
232 surface or mixed into the top three to six inches of soil (Sanders and Addo 1993). Suggested rates of  
233 application are reported as 0.75 pounds of lignin sulfonate per square yard of surface per inch of depth  
234 treated, or more than 10,000 pounds of lignin sulfonate per acre (Harmon 1957). Alternatively, lignin  
235 sulfonate solution containing 25 percent solids are applied at a rate of 7,260 gallons per acre (Parametrix,  
236 Inc. 2003). The use of the petitioned substance is for dust control in the context of a plant or soil

237 amendment, such as in a blended or specialty fertilizer applied to organic crops. Use rates in a fertilizer  
 238 would be much lower, with the lignin component typically comprising 10 percent or less of a fertilizer  
 239 formulation and applied at rates of approximately 50–200 pounds of lignin sulfonate per acre (OMRI 2020).  
 240

241 *Lignins used for chelation*

242 Lignins, particularly lignin sulfonates, are used to chelate micronutrients in fertilizer formulations  
 243 (Agrawal, Kaushik and Biswas 2014) and form more stable, yet still soluble, complexes that improve plant  
 244 micronutrient availability. Forms of lignin sulfonates used as chelating agents include ammonium lignin  
 245 sulfonate (Wallace and Ashcroft 1956) and other salts of lignin sulfonic acid (Keenportz 1995) resulting  
 246 from the sulfite pulping process, such calcium, magnesium, and sodium lignin sulfonate (Bennett 1955).  
 247 These four forms of lignin sulfonates are the most commonly used in organic fertilizer applications (OMRI  
 248 2020). There are many other salts of lignin sulfonic acid, including ammonium calcium lignin sulfonate,  
 249 potassium lignin sulfonate, and zinc lignin sulfonate. Lignins can also chelate heavy metal ions, and as  
 250 such are used in soil remediation (Garcia-Valls and Hatton 2003). Most literature on the use of lignins as  
 251 chelating agents covers only lignin sulfonates; one report does describe the complexation (i.e., chelation) of  
 252 iron cations by lignin anions in soda black liquor (Ghatak 2013). Because the lignins in this report share  
 253 common properties – such as being large phenolic polymers of complex structure with a diversity of  
 254 polymeric unit sizes and an abundance of functional groups – it is expected that other lignin products  
 255 function in a chelating capacity similar to what is seen with lignin sulfonates. The mechanism for lignin  
 256 chelation is discussed below under *Action of the Substance*.  
 257

258 *Other uses for lignins*

259 The primary use for lignin byproducts of the pulping industry is onsite at paper mills as boiler fuel  
 260 (Agrawal, Kaushik and Biswas 2014). Lignins can also be used as a soil additive to reinforce soil structure  
 261 in engineering projects. One study reported the use of sulfur-free lignin to stabilize soil under different  
 262 free-thaw cycles and found that it improved frost resistance (Liu et al 2020). Various lignins have been  
 263 commercialized for use in the formulation of dispersants, adsorbents, adhesives, surfactants,  
 264 nanomaterials, and as starting material for bulk chemicals (Peretti, Barton and Teixeira Mendonca 2016).  
 265 Lignin can be used in engineering plastics such as phenolformaldehyde resins, polyolefin films, and  
 266 polyurethane foams (Košíková and Gregorová 2005). Lignin has also been increasingly employed in  
 267 composites as a reinforcement agent (Liu, et al. 2020). Lignins can be converted into activated carbon,  
 268 vanillin, guaiacol, and carbon fiber (Agrawal, Kaushik and Biswas 2014). Organosolv lignins can be used in  
 269 automotive brakes, biodispersants, and epoxy resins for printed circuit boards (Gallezot 2011). Lignins  
 270 have also been identified as antioxidants (Espinoza-Acosta, et al. 2016; Serrano, et al. 2012; Sun et al., 2018).  
 271 It should be noted that, in many applications, lignins are chemically modified beyond what pulping  
 272 extraction produces; these applications are outside the scope of this report.  
 273

274 **Approved Legal Uses of the Substance:**

275 40 CFR 180.910 lists inert ingredients used pre- and post-harvest that are exempt from the requirement of a  
 276 tolerance. Table 3 below lists the exemptions for lignin and alkali lignin.  
 277

278 **Table 3. Exemptions from the requirement of a tolerance (excerpt from 40 CFR 180.910)**

Lignin (CAS Reg. No. 9005-53-2)	Surfactant, related adjuvants of surfactants
Lignin, alkali (CAS Reg. No. 8068-05-1)	Do*

279 \*A value of "Do" is the CFR representation of "ditto," which refers to uses listed immediately preceding  
 280 that entry in the CFR.  
 281

282 Organosolv lignin (CAS No. 8068-03-9) is subject to reporting at 40 CFR 721.5460, as a significant new use  
 283 chemical substance, identified as any manufacture, processing, or use of the substance with a number  
 284 average molecular weight less than 700 daltons.  
 285

286 See the 2011 Lignin Sulfonate Technical Report for approved legal uses of lignin sulfonates (USDA NOP  
 287 2011).  
 288

**Action of the Substance:**

In nature, lignin helps reinforce plant structures by binding cellulose fibers in wood, thereby helping to stabilize plants against various stresses (Košíková and Gregorová 2005). The same mechanism is seen with extracted lignins in various binding applications, with the binding being dictated by the structural characteristics of any particular lignin extract (Peretti, Barton and Teixeira Mendonca 2016).

Lignins contain reactive groups that allow for many different types of chemical bonding. As aromatic polymers with complex chemical structures, lignins contain aromatic rings with diverse functional groups including hydroxyl, methoxyl, carboxylic, phenolic, sulfonic, and some terminal aldehyde groups (Agrawal, Kaushik and Biswas 2014; Espinoza-Acosta, et al. 2016). Schreiber et al. (2012) noted that lignin's oxygen-containing radicals facilitate crosslinking reactions.

Liu et al. (2020) applied a sulfur-free lignin derived from corn-based ethanol to soil and observed a redistribution of soil particle sizes, with more particles of larger size after the addition of the lignin. They also recorded increases in the water content needed to turn the soil into a semi-solid state (i.e., its plastic limit) and the water content needed to turn the soil into viscous liquid (i.e., its liquid limit). These results suggested increased soil stabilization, and the authors attributed this action to the physical binding of soil particles with lignin into larger aggregates, which also increases soil aeration. They also believed that the low solubility of the lignin could be contributing to the increased water holding capacity observed (Kucera and Sawyer 2015). These binding characteristics, which result in larger particle sizes and greater water holding capacity, also reduce the formation of air-borne dust particles during fertilizer pelletizing and granulating processes.

Lignin's ability to form chemical bonds is also related to the action of chelation, in which a substance binds at more than one point of connection with the chelating agent. Because lignins are large polymers with many polar binding sites, their ability to chelate metal ions and plant nutrients is well documented (Calvo-Flores, et al. 2015). As noted in *Specific Uses of the Substance*, there is little information on the use of other lignins besides lignin sulfonates chelating metals. However, one group of scientists studied the structure of guaiacol aromatic rings linked by  $\beta$ -O-4 linkages—bonds that are one of the most characteristic linkages in lignins and lignin derivatives—complexed with a series of alkali metal cations (e.g., Li<sup>+</sup>, Na<sup>+</sup>, K) using mass spectrometry. Their research found evidence suggesting that the mechanism of complexation was oxygen atoms from multiple  $\beta$ -O-4 linkages involved in chelating the metal (DeBlase, et al. 2016). Additionally, Garcia-Valls and Hatton (2003) report that low molecular weight units in lignin, which contain aromatic rings with certain constituents, complex heavy metal ions. As noted under *Properties of the Substance*, lignins with varying degrees of polydispersity are likely to have some proportion of these low molecular weight units. Thus, it can be expected that other lignin derivatives besides lignin sulfonates would also possess chelating capacities.

**Combinations of the Substance:**

Different lignins contain different residues of substances used in the various pulping processes herein described. Additionally, anthraquinone (AQ) and soluble anthraquinone (SAQ) are substances that have been cited as pulping additives used in the soda lignin extraction process to reduce the degradation of carbohydrates (Biermann 1996). AQ is a polycyclic aromatic hydrocarbon derived from anthracene or phthalic anhydride (NIH 2020) and is not further considered within the scope of lignins addressed in this technical report.

Garcia-Valls and Hatton (2003) described how lignin obtained as a byproduct from plant biomass conversion can be chemically modified to increase its number of active carboxyl, amine, and hydroxyl sites available for complexation with metal ions. This is accomplished through processes such as oxidation with alkaline nitrobenzene or permanganate, acidolysis, hydrogenolysis, or thioacetolysis. This modification releases low molecular weight units containing aromatic rings. However, lignins that are chemically modified subsequent to or beyond the pulping and its associated extraction steps are not considered within the scope of this report, with the exception of lignin sulfonates produced from kraft lignins, as described below under *Evaluation Question #2*.



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<b>Status</b>
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**Historic Use:**

Soda pulping was first invented in England in 1851 using sodium hydroxide as the cooking chemical (Biermann 1996). In the late 1870s, German manufacturers added sodium sulfate to regenerate the sodium hydroxide in the soda pulping process. In doing so, they discovered a much more efficient pulping process with faster delignification, greater selectivity in dissolving the lignin, and less degradation of the cellulose fibers (NC State 2020; Biermann 1996). This led to the development of the kraft process in 1879, which was put into commercial practice in 1890. Most U.S. mills that had used the soda pulping method switched to the kraft process once it was invented (Biermann 1996).

In the 1950s, the U.S. Forest Products Laboratory studied cold soda pulping of hardwoods, where wood chips were soaked in cold sodium hydroxide (5–15 percent); this process was eclipsed by other pulping methods in subsequent decades (Biermann 1996). Currently, soda pulping is reported to have limited use for easily pulped materials such as straw and some hardwoods and is not a predominant process for wood pulping.

Organosolv pulping was developed in the 1990s as an alternative to conventional chemical delignification; however, it has not become economically viable for large-scale operations (Peretti, Barton and Teixeira Mendonca 2016).

**Organic Foods Production Act, USDA Final Rule:**

The petitioned substance is related to lignin sulfonate, which is listed at 7 CFR 205.601(j)(4).

**International**

A survey of regulations and guidelines for organic production from a number of countries and international organizations indicates that the use of sodium carbonate lignin is not generally permitted. The summary below provides international context for the use of sodium carbonate lignin and other lignins in organic crop production.

*Canadian General Standards Board Permitted Substances List. CAN/CGSB-32.311*

Sodium carbonate lignin is not included on PSL Table 4.2 (Soil amendments and crop nutrition) or 4.3 (Crop production aids and materials). Table 4.3 lists “Lignin sulphonates” as “a chelating agent, as a formulant ingredient and as a dust suppressant.” This entry specifically prohibits “ammonium lignosulphonate,” suggesting that “lignin sulphonate” is not intended to be read as a broadly inclusive term for any form of lignin sulfonate.

PSL entries for “Chelates” and “Dust suppressants” on both Tables 4.2 and 4.3, and for “Calcium lignin sulphonate” on Table 4.3, refer to the “Lignin sulphonates” entry on Table 4.3.

*CODEX Alimentarius Commission, Guidelines for the Production, Processing, Labelling and Marketing of Organically Produced Foods (GL 32-1999)*

Sodium carbonate lignin is not included in GL 32-1999 Annex 2 Table 1 (Substances for Use in Soil Fertilizing and Conditioning) or Table 2 (Substances for Plant Pest and Disease Control).

*European Economic Community (EEC) Council Regulation, EC No. 834/2007 and 889/2008*

Sodium carbonate lignin is not included in EC No. 834/2007 Annex I (Fertilisers and soil conditioners referred to in Article 3(1)) nor in Annex II (Pesticides – plant protection products referred to in Article 5(1)). Any reference to materials of wood- or tree-origin specifies that the material must not be chemically treated after felling.

397 *Japan Agricultural Standard (JAS) for Organic Production*

398 Sodium carbonate lignin does not appear in Notification No. 443 Japanese Agricultural Standard for  
399 Organic Plants Partial Revision March 27, 2017. "Lignin sulfonic acid" is allowed in the same Notification  
400 as a 'granulating agent and anticaking agent for fertilizer' 'in case of difficulty to manufacture granulating  
401 agent and anticaking agent from these substances [natural sources]' on Attached Table 1 Fertilizers and soil  
402 improvement substances.

403

404 *IFOAM-Organic International*

405 Sodium carbonate lignin is not included in the IFOAM Norms Appendix 2: Fertilizers and Soil  
406 Conditioners nor in Appendix 3: Crop Protectants and Growth Regulators. "Wood, bark, sawdust, wood  
407 shavings, wood ash, wood charcoal" are included, with their Conditions for Use being "only if not  
408 chemically treated."

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410

### 411 Evaluation Questions for Substances to be used in Organic Crop or Livestock Production

412

413 **Evaluation Question #2: Describe the most prevalent processes used to manufacture or formulate the**  
414 **petitioned substance. Further, describe any chemical change that may occur during manufacture or**  
415 **formulation of the petitioned substance when this substance is extracted from naturally occurring plant,**  
416 **animal, or mineral sources (7 U.S.C. § 6502 (21)).**

417

418 The processes used to derive commercial lignins from plant matter characterize the different lignin  
419 products, since the delignification method influences the final lignin's structure (Serrano, et al. 2012). All of  
420 the processes covered in this report have as their primary purpose the delignification of wood or other  
421 plant material in order to liberate the plant cellulose for paper fiber production. Other manufacturing  
422 processes not covered in this report include biofuel production resulting in lignins as byproducts, as well  
423 as other processes aimed at further modification of extracted lignins for added functionality. The four main  
424 industrial processes that isolate high-purity lignin are the soda, sulfite, kraft, and organosolv processes  
425 (Strassberger, Tanase and Rothenberg 2014); these are discussed below in Table 4 along with other more  
426 minor processes.

427

428 *Soda pulping*

429 The petition describes the process used to manufacture sodium carbonate lignin both as soda ash pulping  
430 and soda pulping. While there is some overlap, the scientific literature predominantly reports the singular  
431 use of sodium hydroxide as the cooking chemical in soda pulping (Espinoza-Acosta, et al. 2016; Calvo-  
432 Flores, et al. 2015; Hellstén, et al. 2013). Hellstén et al. (2013) performed soda extraction on hardwoods and  
433 softwoods at cooking temperatures of 165°F and 170°F, respectively, in a liquor-to-wood ratio of 4:1 and an  
434 effective alkali of 5.5mol/kg. Another study used an extraction solution of 7.5 percent weight per weight  
435 (w/w) sodium hydroxide with a solid-to-liquid ratio of 1:18 at 90°C for 90 minutes on apple wood  
436 (Serrano, et al. 2012). Lignin recovery in soda pulping may be done by acid precipitation (sulfuric acid)  
437 (Serrano, et al. 2012), maturation and filtration (Strassberger, Tanase and Rothenberg 2014), or  
438 centrifugation (Serrano, et al. 2012). The resulting lignins are sulfur-free.

439

440 One report described the use of alkaline solutions, typically sodium hydroxide, under conditions similar to  
441 kraft pulping (see below) but without the inclusion of hydrogen sulfide anions (Espinoza-Acosta, et al.  
442 2016). In this process, the native lignin network undergoes hydrolytic cleavage, thereby extracting the  
443 lignins into the soda black liquor. The authors describe the process of soda lignin recovery from the black  
444 liquor using acid precipitation with various mineral acids, heating, and filtration (Espinoza-Acosta, et al.  
445 2016). Precipitation may also be done with carbon dioxide (Hellstén, et al. 2013); however, both methods of  
446 precipitation have drawbacks: the formation of colloids which impedes subsequent filtration, the low  
447 efficiency of lignin yield with carbon dioxide, and the high cost to incorporate mineral acid precipitation in  
448 pulp mills (Hellstén, et al. 2013).

449

450 The soda process is commonly employed in the pulping of herbaceous feedstocks such as wheat straw,  
451 hemp, and bagasse. Calvo-Flores et al. (2015) noted that the characteristic of the feedstock determines, in

452 part, the ease with which lignin can be obtained by the soda process. Wood lignin requires more aggressive  
453 treatment (5 percent NaOH at 130–170°C), while grass lignins can be isolated by mild alkaline treatments  
454 even at room temperature (Calvo-Flores, et al. 2015).

455

#### 456 *Soda ash pulping*

457 Soda ash pulping can be considered a type of soda pulping. Sodium carbonate lignin, as described in the  
458 petition, results from a process wherein wood chips are subjected to a solution of either sodium carbonate  
459 (6–8 percent solution) or a combination of sodium carbonate (50–85 percent) and sodium hydroxide (15–  
460 50 percent) and cooked in steam under 1100 kPa pressure for approximately 14 minutes (Legnochem 2019).  
461 The cooked woodchips are then mechanically pressed to separate the spent cooking liquor, also known as  
462 the black soda liquor, which is then evaporated to a solution containing 45–55 percent solids, or evaporated  
463 until a dry powder is obtained (Legnochem 2019). This process is consistent with the patent referenced in  
464 the petition, which also notes an ideal pulping solution of 20 percent sodium hydroxide and 80 percent  
465 sodium carbonate, and further specifies a cooking temperature of 375°F (Dillard, Gilmer and Kennedy  
466 1974).

467

468 Another study evaluated differing mixtures of sodium carbonate and sodium hydroxide as cooking  
469 chemicals for pulping wheat straw (Marin, et al. 2017). Their process was to dissolve analytical grade  
470 sodium hydroxide and sodium carbonate in tap water, heat for 30 minutes and cook wheat straw in the  
471 liquor at a ratio of 5:1 liquid to solids for 60 minutes at 150°C, 160°C, or 170°C. The alkali charge was also  
472 varied by chemical concentrations of 16, 18, or 20 percent sodium hydroxide and mass ratio of sodium  
473 carbonate to sodium hydroxide of 1, 5, and 9. The study found that increased sodium hydroxide  
474 concentration and increased temperature promoted delignification, but also increased polysaccharide  
475 degradation, which was undesirable in terms of pulp yield (Marin, et al. 2017).

476

#### 477 *Organosolv pulping*

478 Organosolv pulping uses organic solvents such as organic acids, alcohols, or ketones to delignify pulp and  
479 may or may not include mineral or organic acid catalysts. This manufacturing process involves separating  
480 lignin from plant biomass via acid-catalyzed solubilization, in which bonds such as  $\alpha$ -aryl ether and aryl  
481 glycerol- $\beta$ -aryl ether bonds in lignin macromolecules are cleaved (Espinoza-Acosta, et al. 2016). One study  
482 reported three organosolv processes (Gonçalves and Schuchardt 1999). The first used a  
483 methanol/water/NaOH/AQ mixture on spruce wood to produce what was called “Organocell lignin.”  
484 The second used 93 percent aqueous acetic acid and 0.3 percent hydrochloric acid on bagasse or eucalyptus  
485 chips under reflux for 2 hours, followed by 80 percent evaporation of the acetic acid from the pulping  
486 liquor and precipitation of the lignin with water, filtering, and drying. The third used an  
487 acetone/water/FeCl<sub>3</sub> extraction of lignin from bagasse (Gonçalves and Schuchardt 1999). Lignins derived  
488 from organosolv methods using acetic acid may be termed “Acetosolv lignin.” Another process using  
489 acetic acid and phosphinic acid observed up to 88 percent delignification of birch chips cooked at 150°C  
490 over 120 minutes, and noted that the rate and structure of delignification followed a function of time (VTT  
491 Technical Research Center 2015). Further, an organosolv extraction of apple wood prunings used  
492 60 percent volume per volume (v/v) ethanol-water solution with a solid-to-liquid ratio of 1:10 at 180°C for  
493 90 minutes in a stirred and pressurized reactor, after solids were filtered from the black liquor and the  
494 lignin precipitated with acid water and dried (Serrano, et al. 2012).

495

#### 496 *Kraft pulping*

497 The most predominant process for pulping wood in paper manufacturing and for obtaining lignin from  
498 lignocellulosic biomass is the kraft process (Ahmad and Pant 2018). This process uses sodium hydroxide  
499 and sodium sulfide to dissolve lignin from wood. Ahmad & Pant (2018) specify the temperature range of  
500 150–180 °C for 2 hours under high pH conditions. This ionizes the phenolic hydroxyl groups, thereby  
501 solubilizing the lignin content of the lignocellulosic biomass. Graymont Ltd. (2020) describes loop systems  
502 that together generate sodium hydroxide in kraft mills. In one loop, sodium hydroxide is pumped into  
503 chambers where it is mixed with wood chips under heat and pressure. The sodium hydroxide attacks the  
504 lignin and in the process is converted to sodium carbonate. This sodium carbonate is then pumped into a  
505 vessel containing calcium hydroxide (slaked lime). These react to form sodium hydroxide and calcium  
506 carbonate. The sodium hydroxide is pumped off for reuse, and the calcium carbonate is transferred to a

507 kiln, heated, and thereby converted to calcium oxide (lime). The calcium oxide is mixed with water and  
 508 again forms calcium hydroxide (slaked lime). The sodium carbonate returning from the pulping process is  
 509 then added to the mixture to form sodium hydroxide, and the whole process begins again (Graymont Ltd.  
 510 2020).

511  
 512 Ahmad & Pant (2018) describe the lignin and hemicellulose that is collected in the black liquor being sent  
 513 to evaporators and concentrated. They also note that the sulfur and caustic content is recycled, and that  
 514 lignins destined for polymer production are precipitated by lowering the pH with carbon after which the  
 515 lignins are filtered and washed. This filtering removes sulfur compounds from the final lignin product.  
 516 Lignosulfonates can also be produced from the precipitated kraft lignin by sulfonation with bisulfate or a  
 517 sulfite compound.

#### 518 519 *Sulfite chemical pulping*

520 Sulfite chemical pulping is the most predominant lignin-producing process, generating lignin sulfonate  
 521 (Strassberger, Tanase and Rothenberg 2014). In this process, wood is cooked in an aqueous solution of  
 522 sulfur dioxide (SO<sub>2</sub>) and a base (calcium, sodium, magnesium, or ammonium) (Espinoza-Acosta, et al.  
 523 2016). The lignin sulfonates that are extracted into the black liquor are further purified to remove excess  
 524 sugars through fermentation of the liquor followed by heating to remove generated alcohol (USDA NOP  
 525 2011), ultrafiltration, chemical removal, or selective precipitation of carbohydrates (Strassberger, Tanase  
 526 and Rothenberg 2014). Acid sulfite pulping, alternatively, uses sulfurous acid and sodium, magnesium,  
 527 calcium, or ammonium bisulfite (USDA NOP 2011).

#### 528 529 *Neutral sulfite semichemical (NSSC) pulping*

530 The soda process is sometimes referred to as a “semi-chemical pulping” process that does not involve the  
 531 use of any sulfur compounds or bleaching chemicals (Legnochem 2019). Another process is referred to as  
 532 “neutral sulfite semichemical pulping” (Dillard, Gilmer and Kennedy 1974). This process uses a liquor of  
 533 sodium sulfite and sodium carbonate in a molar ration of 7:1–3:1, with optional addition of buffers such as  
 534 sodium bicarbonate or sodium hydroxide. Alternatively, this process may involve sodium sulfite and  
 535 sodium bicarbonate in a ratio 5:1 at 14 percent chemical concentration, cooking wood at 170°C and 690 kPa  
 536 for 2–3 hours. However, this process results in a liquor containing hydrogen sulfide and sulfur dioxide and  
 537 is thus less desirable due to the malodor and corrosiveness (Dillard, Gilmer and Kennedy 1974).

#### 538 539 *Steam explosion delignification*

540 Autohydrolysis and steam-explosion are treatments wherein the plant biomass is separated into a liquid  
 541 stream composed primarily of hemicelluloses, oligomers, and a small fraction of water-soluble lignin  
 542 (Peretti, Barton and Teixeira Mendonca 2016). Such process parameters have been reported as 150–220°C  
 543 (Peretti, Barton and Teixeira Mendonca 2016) and 160 °C for 15–210 minutes in a water-to-wood ratio of 4:1  
 544 (Lu, et al. 2012). This is sometimes done as a pre-treatment prior to soda pulping (Liu, et al. 2020; Lu, et al.  
 545 2012), but it is not effective for obtaining lignins from wood on its own. One report of a hot-water pre-  
 546 treatment prior to soda pulping found that less than 6 percent lignin (on total wood mass) from virgin  
 547 woodchips was dissolved in the extraction liquor with hot water extraction of aspen woodchips. Typical  
 548 pulping extraction yields are around 80 percent of lignin from wood (Lu, et al. 2012).

#### 549 550 *Enzymatic hydrolysis*

551 Lignins can also be obtained by successive treatment with selective enzymes that hydrolyze carbohydrates;  
 552 however, not all carbohydrates can be removed enzymatically to obtain the same level of purity obtained  
 553 by the other methods typically used for milled wood (Agrawal, Kaushik and Biswas 2014).

554  
 555 **Table 4. Summary of reviewed processes for lignin production. See *Evaluation Question #2* text for  
 556 references.**

<u>Lignin type</u>	<u>Reagents</u>	<u>Reagent concentration</u>	<u>Time</u>	<u>Temperature</u>	<u>Recovery Steps</u>
<b>Soda pulping</b>	Sodium hydroxide	Alkali of 5.5mol/kg	Not reported	165°C (hardwood) and 170°C (softwood)	Not reported

	Sodium hydroxide	7.5% (w/w)	90 minutes	90°C	Acid preparation; maturation and filtration; centrifugation
<b>Soda ash pulping</b>	Sodium carbonate and sodium hydroxide	50-85% carbonate and 15-50% hydroxide	14 minutes	Cooked in steam under 1100 kPa pressure	Mechanical pressing and evaporation
	Sodium carbonate and sodium hydroxide	20% hydroxide and 80% carbonate	Not reported	375°F	Not reported
	Sodium hydroxide and sodium carbonate	16, 18, 20%	60 minutes	150-170°C	Not reported
<b>Organosolv pulping</b>	Methanol/ water/ NaOH/ AQ	Not reported	Not reported	Not reported	Not reported
	Aqueous acetic acid; hydrochloric acid	90%; 0.3%	2 hours	Not reported	Evaporation
	Acetic acid and phosphinic acid	Not reported	120 minutes	150°C	Not reported
	Ethanol and water	60% (v/v)	90 minutes	180°C	Filtration and precipitation
<b>Kraft pulping</b>	Sodium hydroxide and sodium sulfide	High pH conditions	2 hours	150-180°C	Precipitation; filtering; washing
<b>Sulfite chemical pulping†</b>	Aqueous solution of sulfur dioxide and a base	Not reported	Not reported	Not reported	Fermentation; ultrafiltration; chemical purification; precipitation
<b>Neutral sulfite semichemical (NSSC) pulping</b>	Sodium sulfite and sodium carbonate	Molar ratio of 7:1-3:1	Not reported	Not reported	Not reported
	Sodium sulfite and sodium bicarbonate	5:1 ratio at 14%	2-3 hours	170°C and 690 kPa	Not reported
<b>Steam explosion delignification</b>	N/A	Water-to-wood ratio of 4:1	15-210 minutes	150-220°C	Not reported

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560

As described above and illustrated in Table 4, the process for obtaining the petitioned substance and the manufacturing process for lignin sulfonate differ principally in the chemicals used to extract the lignins. Sulfite pulping, which produces lignin sulfonates, uses sulfur dioxide and a base, whereas soda ash

561 pulping uses sodium carbonate and sodium hydroxide as the reagents. The literature notes the difference  
562 between the resulting lignins primarily in terms of polymer size and sulfur content.

563  
564 **Evaluation Question #4: Describe the persistence or concentration of the petitioned substance and/or its**  
565 **by-products in the environment (7 U.S.C. § 6518 (m) (2)).**

566  
567 Lignins are a naturally occurring component of wood and other plant biomass and are the second largest  
568 source of natural organic polymers after cellulose (Ponnusamy, et al. 2019; Liu, et al. 2020). As such, they  
569 play an important role in soil carbon cycles, particularly for carbon storage (Thevenot, Dignac and Rumpel  
570 2010). They have historically been considered recalcitrant, or resistant to biodegradation. Only a few  
571 organisms are able to degrade lignins' complex structure, including *Streptomyces* sp., *Nocardia* sp., and  
572 basidiomycete brown-rot and white-rot fungi. Of these, most can alter lignin structure, but only white-rot  
573 basidiomycetes are able to completely mineralize lignin molecules (Thevenot, Dignac and Rumpel 2010;  
574 Sierra-Alvarez and Lettinga 1991). There may also be anaerobic organisms and abiotic factors that can  
575 contribute to lignin degradation in the soil.

576  
577 Degradation rates in the field have been found to be extremely variable, but Thevenot et al. (2010)  
578 summarized them as being between 48–87 percent over 5 years. These rates may not be slower than those  
579 of bulk soil organic carbon and may even be faster (Thevenot, Dignac and Rumpel 2010). Degradation rates  
580 are influenced by many factors in the environment, including climate, soil type, pH, moisture level, and  
581 management, as well as soil microbial communities. For lignin derivatives, polymer size is also an  
582 important factor in degradability. Lignin monomeric model compounds representative of those derived  
583 from the soda process are reportedly degradable under anaerobic conditions. Kraft lignins subjected to  
584 mixed methanogenic (anaerobic) cultures at moderate to high temperatures showed degradation, but only  
585 of low molecular weight material (i.e., less than 600 daltons). Native lignin and higher molecular weight  
586 synthetic lignins are highly resistant to anaerobic degradation. Aerobic bacteria are also reported to  
587 metabolize low molecular weight lignin fractions (Sierra-Alvarez and Lettinga 1991).

588  
589 One study applied acid-precipitated kraft lignin to soil samples in order to evaluate soil properties,  
590 including the potential for buildup of sodium in the soil (Xiao, Bolton and Pan 2007). The lignin was  
591 applied at rates of 1.67 and 3.34 g C kg<sup>-1</sup> soil and incubated for 8 weeks. The results showed an increase in  
592 electrical conductivity with increasing lignin application; however, the authors concluded that the increase  
593 was small enough that lignin application at these rates would not pose concerns for soil salinity. Stiggson  
594 and Lindstrom (2007) found that washing lignin that had been precipitated from alkali kraft black liquor  
595 with carbon dioxide, in a solution containing magnesium or calcium effectively replaced many of the  
596 sodium ions with the magnesium or calcium. Beyond these reports, there is little information in the  
597 literature regarding the sodium content of sodium carbonate lignin, soda lignin, or other alkali lignins used  
598 in dust suppression and their potential to affect the buildup of sodium in the soil.

599  
600 The study by Xiao, Bolton and Pan (2007) suggested the potential benefit of lignin as a soil amendment,  
601 reporting that lignin in the soil acts as an important precursor in the formation of humic substances. Their  
602 study found an increase in soil organic carbon and total nitrogen with application of the lignin, as well as  
603 increased macroaggregate formation and stability.

604  
605 **Evaluation Question #5: Describe the toxicity and mode of action of the substance and of its**  
606 **breakdown products and any contaminants. Describe the persistence and areas of concentration in the**  
607 **environment of the substance and its breakdown products (7 U.S.C. § 6518 (m) (2)).**

608  
609 Paper mill wastewater streams have been identified as a significant environmental challenge (Mandeep  
610 and Shulka 2020). These waste streams contain lignocellulosic materials with various organic and inorganic  
611 polluting complexes that result not just from pulping but also bleaching processes, e.g., chlorophenols  
612 (Mandeep and Shulka 2020). Lignin components in this waste stream have been studied for toxicity; one  
613 study attributed pulping effluent toxicity in the form of enzymatic inhibition to hydrophilic, higher-  
614 molecular weight lignin fractions using reverse electron transport (Pessala, et al. 2004). The authors of this  
615 study found lignins in pulp wastewater to be toxic to the aquatic organisms *D. magna* and *V. fischeri*

616 (Pessala, et al. 2004). These findings are related to lignins in wood pulping effluents. Removing lignins,  
617 including the petitioned substance, from pulping wastewaters to convert them into value-added products  
618 (valorization) such as dust suppressants, fertilizer chelating agents, and other products could therefore  
619 have some environmental benefits.

620  
621 In contrast, a summary report on the advantages and disadvantages of lignin valorization concluded, upon  
622 review of numerous environmental studies over the last 30 years, that lignin sulfonates are nontoxic at  
623 concentrations above those used, and that surrounding vegetation is not affected by the application of  
624 lignosulfonates (Strassberger, Tanase and Rothenberg 2014). One study found higher onset of degradation  
625 for sulfur-free lignin (trade name Polybind) than for a calcium lignin sulfonate (Sahoo, et al. 2011).

626  
627 Espinoza-Acosta et al. (2016) investigated lignin-derived polyphenols for their antioxidant capabilities,  
628 indicating their advantage over more conventional synthetic antioxidants due to their biodegradability and  
629 low toxicity.

630  
631 **Evaluation Question #6: Describe any environmental contamination that could result from the**  
632 **petitioned substance's manufacture, use, misuse, or disposal (7 U.S.C. § 6518 (m) (3)).**

633  
634 The paper industry creates a significant environmental burden in the form of emissions (U.S. EPA 2018),  
635 effluent, and solid waste (Mandeep and Shulka 2020). Paper mill effluents from processes such as kraft  
636 pulping, neutral sulfite semichemical pulping, and bleaching are reported to be poorly degradable (Sierra-  
637 Alvarez and Lettinga 1991). Removal of lignins from the effluents is desirable, and alkali lignin is readily  
638 separated via acid precipitation and filtration (Sierra-Alvarez and Lettinga 1991). The solid waste lignins  
639 recovered from these and other pulping effluents are primarily used as a fuel in the pulping plant.  
640 However, this use of the lignin byproduct has been reported to have low efficiency as it is a lower-grade  
641 fuel (Strassberger, Tanase and Rothenberg 2014), and the burning of lignins in these fuels adds to  
642 atmospheric pollution through the generation of guaiacol (2-methoxyphenol) (Pflieger and Kroflič 2017).  
643 Thus, diverting pulp waste away from use as fuel reduces the air pollution generated from the wood  
644 pulping process. Mandeep & Shulka (2020) did report on the environmental benefits of diverting pulp  
645 wastes, including lignin, from the waste stream and into biorefinery applications, which included low  
646 emissions and relatively high efficiency. Garcia-Valls & Hatton (2003) similarly suggested that the  
647 environmental burden posed by paper pulping waste lignins can be largely addressed through utilization  
648 of the lignin materials in value-added products. One study reported the soda process to have the lowest  
649 potential environmental impact of four pulping processes considered, followed by the sulfite, kraft and  
650 organosolv processes, as measured by a waste reduction algorithm developed by the EPA's National Risk  
651 Management Research Laboratory (Carvajal, Gómez and Cardona 2016). None of the literature reviewed  
652 for this report suggested that the use of lignins as petitioned would result in environmental contamination  
653 of organic systems.

654  
655 **Evaluation Question #8: Describe any effects of the petitioned substance on biological or chemical**  
656 **interactions in the agro-ecosystem, including physiological effects on soil organisms (including the salt**  
657 **index and solubility of the soil), crops, and livestock (7 U.S.C. § 6518 (m) (5)).**

658  
659 Liu et al. (2020) investigated soil stabilization with sulfur-free lignin obtained as a byproduct of ethanol  
660 production and found a slight decrease in soil pH with application of the lignin, but it was still comparable  
661 to the pH of natural soil. They therefore concluded that the sulfur-free lignin did not significantly change  
662 the soil pH, despite previous studies finding alkalization of soils treated with lignin sulfonate byproducts  
663 of the paper industry. Xiao et al. (2007) found that acid-precipitated kraft lignin significantly decreased soil  
664 pH over 8 weeks of incubation and may therefore act as an acidifying agent in the soil. The authors  
665 postulated that the acidification observed was likely due to the lignin having been precipitated with acid,  
666 as well as nitrification of ammonia nitrogen in the lignin byproduct and mineralization of organic matter in  
667 the soil. Liu et al. (2020) also reported a slight increase in the soil cation exchange capacity with addition of  
668 the sulfur-free lignin at rates of 3–15 percent lignin applied to soil samples.

669

670 Different types of lignin phenols can be inhibitory to some fungal species (Guiraud, et al. 1995); however,  
671 they may also stimulate microbial activity depending on the amount of lignin applied (Xiao, Bolton and  
672 Pan 2007). In Xiao et al.'s (2007) study, application of a lignin amendment at rates up to 6.12 g kg<sup>-1</sup> soil  
673 appeared to increase soil microbial activity, observed as increased soil nitrification. They also observed an  
674 increase in the proportion of larger soil aggregates. While this may be due to the binding nature of the  
675 lignins applied, the authors also note the potential contribution of hyphal growth of fungi responsible for  
676 lignin degradation (Xiao, Bolton and Pan 2007).

677  
678 The use of the petitioned substance is for dust control in the context of a plant or soil amendment, and as  
679 such is applied to the soil at relatively low rates. This consideration, in combination with the potential  
680 effects of lignin to increase soil aggregate size and water retention (Kucera and Sawyer 2015) and enhance  
681 microbial activity (Xiao, Bolton and Pan 2007), suggest that its use as petitioned is likely to be benign and  
682 may even be ecologically beneficial.

683  
684 **Evaluation Question #9: Discuss and summarize findings on whether the use of the petitioned**  
685 **substance may be harmful to the environment (7 U.S.C. § 6517 (c) (1) (A) (i) and 7 U.S.C. § 6517 (c) (2) (A)**  
686 **(i)).**

687  
688 A review of scientific literature for this technical report did not uncover any reports of environmental harm  
689 resulting from the use of lignins, including sodium carbonate lignin, as petitioned.

690  
691 Some environmental benefits of lignins, in addition to those already mentioned, include dust suppression  
692 and slow release of chelated micronutrients. Lignins used in these applications are not applied directly to  
693 the soil, but to fertilizers and soil amendments during their manufacture. Fertilizer dust inhalation can pose  
694 health concerns, and fertilizer dust can contribute to air pollution and surface water contamination (Kucera  
695 and Sawyer 2015). Micronutrients in the soil can be unavailable to plants in certain pH ranges. However,  
696 adding these micronutrients in chelated form keeps them bioavailable, as the organic macromolecule of  
697 lignin envelops the micronutrient and prevents it binding with other inorganic elements in the soil that  
698 would inhibit its uptake by plants (Brady and Weil 2002).

699  
700 **Evaluation Question #11: Describe all natural (non-synthetic) substances or products which may be**  
701 **used in place of a petitioned substance (7 U.S.C. § 6517 (c) (1) (A) (ii)). Provide a list of allowed**  
702 **substances that may be used in place of the petitioned substance (7 U.S.C. § 6518 (m) (6)).**

703  
704 The petitioned substance is proposed for use as a dust suppressant. At the time of this report, OMRI has  
705 11 products listed in the category of "Dust Suppressants" and another 29 in the category of "Lignin  
706 Sulfonates" (OMRI 2020). Products listed in the "Dust Suppressants" category may contain dust-  
707 suppressing agents such as vegetable oils, magnesium or calcium chloride solutions, glycerin, natural fatty  
708 acids, or synthetic lignin sulfonates. Wax emulsions or combinations of oil and wax emulsions have also  
709 been reported for dust control; however, their effectiveness over time may decrease as oils volatilize or  
710 become adsorbed into fertilizer particles (Kucera and Sawyer 2015). Waxes have proven difficult to handle  
711 unless at temperatures where they are melted and can absorb into fertilizer particles. Both waxes and oils  
712 also have limited binding capacity (Kucera and Sawyer 2015).

713  
714 Other proposed materials for use in dust control methods include solutions containing molasses, or simple  
715 application of water. These methods, however, can increase fertilizer caking and their binding properties  
716 are lost with drying, thus rendering them ineffective for longer-term dust control (Kucera and Sawyer  
717 2015).

718  
719 Other nonsynthetic fibrous materials have been suggested for use as binders to control dust during  
720 fertilizer granulation. These include wood flour, peat moss, compost, manure, cotton, straw, brewers  
721 condensed solubles, beet syrup, whey starch, soy solubles, corn cob, rice hulls, peanut hulls, ground wheat  
722 straw flour, wheat flour, soy flour, cellulose derivatives, seed meal, feather meal, soy meal, and humic acid  
723 (Kucera and Sawyer 2015). One study reported the use of anaerobic digestate filtrate and limestone used in  
724 the granulation of fertilizer. These materials were mixed and dried, yielding granules of acceptable



725 strength (Mangwandi, et al. 2013). Clay additives have also been reported to have consistent and  
726 predictable performance in dust suppression applications (Bolander 1997).

727  
728 Nonsynthetic alternative chelating agents to lignins or lignin sulfonates include amino acids (Hsu 1996)  
729 and citric acid (USDA NOP 2011).

730  
731 **Evaluation Question #12: Describe any alternative practices that would make the use of the petitioned**  
732 **substance unnecessary (7 U.S.C. § 6518 (m) (6)).**

733  
734 Agricultural dusts have been reported as a health hazard to farm workers, causing acute and chronic  
735 respiratory problems (Kirkorn and Vincent 2000). These effects can be mitigated in some circumstances by  
736 engineering controls such as misting and mechanical ventilation systems inside farm equipment. Personal  
737 protective equipment in the form of respiratory devices are another option, but their use in practice is  
738 limited in part because they are hot and uncomfortable to wear, and because workers may not be aware of  
739 the risks of agricultural dusts and therefore not think to use the devices (Kirkorn and Vincent 2000).

740  
741 More effective, alternative methods of dust suppression are reported in the 2011 Lignin Sulfonate Technical  
742 Report, and include management practices such as maintaining vegetative cover in non-traffic areas,  
743 mulching, application of stone or gravel to disturbed roads, and air current barriers such as trees. The  
744 report notes that the choice of dust control method may be influenced by different costs, available  
745 resources, and extent of dust control needed (USDA NOP 2011).

746  
747

#### Report Authorship

748  
749  
750 The following individuals were involved in research, data collection, writing, editing, and/or final  
751 approval of this report:

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758 All individuals are in compliance with Federal Acquisition Regulations (FAR) Subpart 3.11 – Preventing  
759 Personal Conflicts of Interest for Contractor Employees Performing Acquisition Functions.

760  
761

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